

**Rational combination platform trial design for children and young adults with Diffuse Midline Glioma:
a report from PNOG**

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Abstract

Diffuse midline glioma (DMG) is a devastating pediatric brain tumor unresponsive to hundreds of clinical trials. Approximately 80% of DMGs harbor H3K27M oncohistones, which reprogram the epigenome to increase the metabolic profile of the tumor cells. We have previously shown preclinical efficacy of targeting both oxidative phosphorylation and glycolysis through treatment with ONC201, which activates the mitochondrial protease ClpP, and paxalisib, which inhibits PI3K/mTOR, respectively. This combination treatment aimed at inducing metabolic distress led to the design of the first DMG-specific platform trial PNOC022 (NCT05009992). Here, we expand on the PNOC022 rationale and discuss various considerations, including liquid biome, microbiome, and genomic biomarkers, quality-of-life endpoints, and novel imaging modalities, such that we offer direction on future clinical trials in DMG.

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Background

Diffuse midline glioma (DMG) is the most lethal childhood cancer, with a median survival of only 10 months.¹ There are 200-300 cases of DMG in the United States annually, each arising in midline structures, such as the spinal cord, pons, and thalamus.^{2,3} Approximately 80% of DMGs harbor somatic missense mutations which substitute methionine for lysine at position 27 of Histone H3.1 or H3.3, known together as the H3K27M mutations.⁴⁻⁷ These H3K27M mutations drive global epigenetic aberrancies including genomic hypomethylation, decreased H3K27 trimethylation (H3K27me₃) and increased H3K27 acetylation (H3K27ac).⁸⁻¹³

H3K27M-mutant DMGs depend on methionine metabolism, with restriction of this amino acid increasing survival of *in vivo* models.¹⁴ To exploit tumor dependency on high metabolic activity, we performed a preclinical investigation of the mitochondrial targeting drugs ONC201 and ONC206, demonstrating that they decrease DMG cell viability *in vitro* and increase survival of *in vivo* models.¹⁵ ClpP activation is achieved by a number of drugs including ADEPs, D9, TR and ONC drugs. ClpP targeting was first achieved as an anti-bacterial strategy using the natural products acyldepsipeptide antibodies (ADEPs) which showed efficacy in ClpP activation.¹⁶ D9 is a small molecule that mimics the natural structure of ClpX, the ClpP chaperon.¹⁷ D9 has been used as a potent ClpP agonist to better understand the function of this mitochondrial protease.¹⁷ TR compounds are a novel series of imipridone that have shown stringent efficacy in ClpP targeting with some reported to be an order of magnetite more effective than ONC compounds in inducing breast tumor cell kill.¹⁸ ONC201, the original imipridone drug, was reported to induce TRAIL apoptotic pathway by inhibiting Akt and ERK phosphorylation of Foxo3a. However, subsequent studies showed ONC201 as ClpP agonist.¹⁶⁻²¹ Caseinolytic peptidase proteolytic subunit (ClpP) protein localizes to the inner mitochondrial membrane and hydrolyses excess or misfolded proteins.^{20,22} ClpP is a key regulator of the mitochondrial stress response, and a target for disruption of mitochondrial metabolic pathways in cancer. ONC201 efficacy has been studied in a number of cancers including glioblastoma, hematological malignancies, and prostate cancer.²³⁻²⁶ Indeed, initial studies in glioblastoma, showed ONC201 efficacy in a patient with K3K27M mutation.REF To validate this, we used preclinical

models of DMG. Our *in vitro* and *in vivo* studies showed that ONC201 and ONC206 allosterically bind to and increase the mitochondrial protease ClpP activity.²⁷ Accordingly, treatment in DMG drives mitochondrial degradation, in turn causing an integrated stress response (ISR) and oxidative phosphorylation (OXPHOS) impairment; interestingly, astrocytic lineage differentiation of tumor cells is also observed.¹⁵ These effects of ONC201 combined with epigenetic downregulation of cell cycle related pathways correlate with improved survival beyond what has been previously seen in this patient population. Specifically, Venneti et al. demonstrated that ONC201 correlated with prolonged overall survival of 21.7 months in patients with newly diagnosed DMG treated after radiation but before disease progression and 9.3 months in patients with progressive disease.²⁸ While ONC206 may have similar properties, the drug remains in early phase, dose escalation studies. This is in contrast to much larger experience with ONC201 and defined recommended dosing. Thus, we aimed to build on these promising findings with single-agent ONC201 through the addition of novel agents on an ONC201 backbone therapy.

However, H3K27M-mutant DMGs are notable for enhanced glycolysis, potentially offering a compensatory mechanism for tumor energetics following diminished OXPHOS activity.²⁹ This metabolic shift is likely mediated by overactivity of the PI3K/mTOR pathway, which indirectly drives glycolysis.³⁰⁻³² The PI3K/mTOR signaling pathway controls multiple cellular processes including metabolism, motility, proliferation, growth, and survival.^{33,34} For example, the highly conserved AMPK protein, is a key energy sensor and homeostasis regulator such as glucose metabolism.³⁵ AMPK promotes glucose uptake by activating PI3K emphasizing the role of PI3K/mTOR in cellular energy metabolism.³⁶ Furthermore, mTOR inhibition leads to suppression of glutathione production in multiple pediatric brain tumors.³⁷ Because glutathione is critically important to mitochondrial function, suppression of glutathione production may increase the efficacy of mitochondrial targeting drugs such as ONC201. The PI3K/mTOR pathway is also activated as a response to ROS accumulation.³⁸ Therefore, we combined ONC201 with GDC-0084 (paxalisib), a clinically available blood-brain barrier (BBB) penetrative PI3K/mTOR inhibitor with promise in atypical rhabdoid teratoid tumor, glioblastoma, and pediatric high-grade glioma (pHGG).³⁹⁻⁴² This allowed us to

investigate the dual inhibition of OXPHOS and glycolysis. We showed that ONC201 plus paxalisib synergizes in altering OXPHOS and glycolysis to induce metabolic distress.⁴³ This led to the design of the first DMG-specific platform trial PNOC022 (NCT05009992).

Here, we report the PNOC022 rationale and design, review the motivating liquid biome and microbiome work leading to the clinical trial, explore genomic biomarkers of response to the treatment, discuss quality-of-life endpoints, consider the integration of novel imaging modalities to monitor and guide treatment across clinical course, and offer direction for the next generation of clinical trials in DMG.

Challenges in clinical trial design for DMGs

The rapid pace of recent discoveries in the field of DMG has invigorated laboratory and translational research efforts. However, the ultimate objective to develop a therapeutic trial that improves outcomes has yet to be fulfilled.

Unique to DMG, the scientific community is engaged in an ongoing debate regarding the choice of endpoints that will most effectively define success or futility of the hypothesis being tested in clinical settings. Numerous challenges further complicate this endeavor, with the rarity of the disease being a significant obstacle. The utilization of imprecise model systems during preliminary laboratory-based investigations and the prioritization of therapies and relevant biomarkers to evaluate in homogeneous patient cohorts pose further difficulties. These considerations must be balanced with the limited resources available for conducting trials. While the ultimate goal is achieving a cure, there are known confounding factors when studying endpoints, including OS endpoints affected by multiple sequential therapies and incomplete knowledge of subtype-specific natural history. More cost-effective, single-arm phase 2 studies require consensus-validated endpoints that use patient-level data from historically and contemporarily treated populations, ensuring uniform characterization. Alternatively, concurrently controlled randomized studies can be employed. Unfortunately, the current "standard of care" for DMG is far from curative, with any meaningful responses primarily attributable to focal radiotherapy. The acquisition of pharmacodynamic biomarkers introduces more

challenges, necessitating multiple tumor tissue samples or less invasive, non-tumor liquid biopsies for longitudinal assessments using validated assays. Given the complexity and challenges posed by DMG, well-conducted clinical research is crucial.

The Food and Drug Administration (FDA) considers concurrently controlled randomized studies as the "gold standard" for determining efficacy. Disease stakeholders for DMG, including patients/families and researchers, would prefer non-randomized studies until a reasonable standard of care is established. An alternative to concurrently controlled trials is utilizing what the FDA calls "external controls." While not always consisting of contemporaneously treated patients, external controls can provide a control arm for confirming endpoints. Traditional historical controls commonly used in phase 2 trials rely on aggregated results and rarely include sufficient individual patient-level data. Thus, these options lack the comprehensiveness or contemporaneity required to serve as adequate external controls. Recent guidance has been published to identify patients who would be reliable for this purpose, with comprehensive longitudinal clinical, biological, and treatment data at the patient-level providing the minimum required information. The intent of such controls is to assume the role of the concurrent treatment control group, theoretically adhering to all eligibility requirements and follow-up assessments as planned for the intervention. The evident advantage would be a significant reduction in the required sample size for determining trial endpoints while minimizing children's exposure to a control arm with an inadequate standard of care. Substantial resources are necessary to develop the enriched cohorts required for clinical trials and regulatory application of external controls. However, the potential benefits include addressing sample size and concurrent control issues and overcoming specific clinical research challenges present in DMG.

Within PNOC022, we aimed to tackle some of these challenges through specific strategies. Firstly, we only advanced combination therapies into the clinical setting after thorough assessments in multiple model systems and laboratories, ensuring a higher confidence level in their potential efficacy. Secondly, we molecularly characterize each patient's tumor to assess outcomes within a subtype-specific context. Additionally, we have incorporated the collection of CSF as a potential surrogate for on-treatment biopsies. Furthermore, our study integrates various correlative studies, including

microbiome investigations and applies contemporary disease-specific, pediatric radiographic assessment criteria through the use of Response Assessment in Pediatric Neuro-Oncology (RAPNO) specific to DMG.

Despite these advancements, endpoints remain a continued challenge in DMG, even with the application of RAPNO guidelines.

Disease cohorts and rationale

PNOC022 encompasses a multi-cohort, multi-arm adaptive trial design. At the outset of trial development, the goal was to create therapy options for patients at all stages of the disease, from initial diagnosis to disease progression. Investigators wanted to a) explore the potential efficacy of the therapeutic approach at each stage of the disease, b) address questions of drug penetration into the tumor with consideration of pre- and post-radiation effects, and c) investigate biomarkers of response/resistance within the tumor tissue. To this end, the trial includes three cohorts – newly diagnosed, pre-radiation (Cohort 1); newly diagnosed, post-radiation (Cohort 2); and recurrent (Cohort 3) (**Figure 1**). Each cohort has pre-surgical and post-surgical arms to allow a treat-biopsy-treat approach for patients who have not yet undergone standard-of-care biopsy/resection and are enrolled before surgery. Assignment into each combination arm is determined through *a priori* randomization at study entry.

The pre-surgical target validation arms require dosing at a specific time point before surgery. This design facilitates the investigation of the pharmacokinetics of drug penetration into the tumor and the pharmacodynamic effects of the drug on tissue. Further in the target validation arms and as means to assess differential drug penetration, we aim to enroll at least five patients each with disease at the following anatomic locations – thalamic, pons, spine. Within Cohorts 1 and 3, investigated drugs are also given concomitantly with upfront or reirradiation. This design allows both safety and efficacy assessment of combination therapy with radiation. After radiation for Cohorts 1 and 3 and at the time of study entry for Cohort 2, patients begin maintenance therapy with combination therapy. This design allows both safety and efficacy assessment of combination therapy with radiation and exploration of added impact of combinatorial strategies. The current

target validation design investigates post-treatment tumor tissue after ONC201 is given on day -1 or days -2 and -1 and paxalisib on day -1. For cohort 1 and 3 patients are then randomized to receive either ONC201 or paxalisib with radiation therapy. Within the maintenance phase for cohort 1 and 3 as well as at study entry for cohort 2, all patients receive the combination therapy of ONC201 and paxalisib. An adaptive trial design allows the incorporation of new combination strategies throughout the life of the trial. By offering a flexible and multi-arm/multi-cohort design, PNOC022 aims to comprehensively address toxicity, efficacy, and biomarker identification for patients with DMG, regardless of disease stage, and across a wide-ranging combination of therapies.

Statistical design (platform design and endpoints)

The trial contains three cohorts based on the stage of the disease and two to three phases within each cohort: target validation, upfront or re-irradiation, and maintenance. Within each disease phase cohort, patients are randomized to one of the study arms or assigned a study arm in the setting of prior exposure to treatment on an alternate arm. The adaptive platform design allows new treatment combination arms to be added and available for additional randomization options during the trial and also provides a mechanism that patients can switch to new cohorts and arms as their disease progresses.

The trial uses the novel Bayesian drug combination platform trial design with adaptive shrinkage (ComPAS) for all cohorts.⁴⁴ ComPAS allows for dropping ineffective drug combinations and adding new combinations to the ongoing clinical pipeline adaptively based on the accumulating trial data. The treatment maintenance combinations are initially formed with ONC201 and paxalisib, with ONC201 as the backbone therapy and paxalisib as the novel agent. For the target validation and radiation phases, assigned patients will receive only single agent, monotherapy according to the study arm to which they were randomized at the study entry. The novel agents and ONC201 combinations will then be given during the maintenance phase. Each combination will undergo two interim analyses, at which point, a decision will be made to stop the arm early for futility, graduate the arm for superiority, or continue the arm to the next interim or final stage. Patients will also be stratified within each arm according to known molecular and clinical prognostic markers of H3 status, *TP53* wildtype versus mutant, and age at diagnosis. The ComPAS design decisions for interim

and final analyses will be made in comparison to the appropriate historical control using a Bayesian hierarchical model. The advantage of this model is the ability to borrow information across arms. There are two settings where borrowing may be advantageous: first, if efficacy (either highly efficacious or not-efficacious) is similar at the interim analysis in the same two arms in Cohorts 1 and 2; second, if efficacy is identical for two arms that only differ by the number of days the drug was given before the target validation phase (i.e., 1 or 2 days).

For Cohorts 1 and 2, the primary objective is to assess the efficacy of the combination therapies based on median progression-free survival at six months (PFS6). PFS6 is defined as the percentage of patients alive and free from progression at six months (26 weeks). The historical control PFS6 rate is 39% based on an ONC201 monotherapy trial (NCT03416530).⁴⁵ The maximum sample size is 33, and the interim analyses will occur when 11 and 22 patients have been enrolled. For Cohort 3, the primary objective is to assess the efficacy of the combination therapies based on median overall survival at seven months (OS7) based on patient-level data from published data.^{46–51} OS7 is defined as the percentage of patients alive at seven months (30 weeks). The historical control OS7 rate is 50% based on published literature. The maximum sample is 42; the interim analyses will occur when 14 and 28 patients have been enrolled.

Correlative studies of PNOC022

Cell free tumor DNA (cf-tDNA)

Over the past decade, liquid biopsy technologies have emerged to monitor the treatment response of various solid tumors on a clinical and molecular level. These technologies involve assessing plasma and cerebrospinal fluid (CSF) for extracted cell-free tumor DNA (cf-tDNA). Recent studies have shown that these liquid biopsy techniques can effectively detect patient-specific tumor variants in both DMG and non-DMG tumors and even correlate or predict tumor growth and response to treatment.^{52–56} We and others have shown the utility of digital droplet PCR (ddPCR) to measure circulating tumor DNA (ctDNA) in H3K27M DMGs.^{53,57–61}

Probing for *H3F3A* (H3.3) K27M mutations, studies showed higher detectable levels of ctDNA in samples of cf-tDNA derived from CSF compared to serum or plasma, as further described below.^{53,57–60} However, the clinical utility of probing H3.3K27M cf-tDNA for therapeutic monitoring of response and resistance in ongoing and prospective trials was unclear. Early clues of the feasibility of clinical assessment of longitudinal ctDNA in DMGs came from a pilot precision trial utilizing upfront biopsies for designing a biopsy-informed therapy (NCT02274987).⁶² This trial reported the detection and monitoring of *H3F3A* and *HIST1H3B* (H3.1) wild-type (WT) and mutant alleles and assessment of mutation allele frequency (MAF) in 11/13 (85%) of subjects at diagnosis, (6/6; 100%) at post-radiation, (7/7; 100%) at treatment, (5/7; 71%) at progression, and (5/5; 100%) at the end of the study. However, the MAF positive detection rate remained very low, hindering further clinical interpretations.

While monitoring MAF is a powerful approach for detecting clinically relevant histone and driver mutations in the pHGG liquid biome, this approach is suitable only for monitoring a limited number of small nucleotide variants (SNVs). It is, therefore, insufficient to detect larger-scale genomic alterations. Capturing intra-tumoral mutational heterogeneity requires comprehensive mutation profiling in ctDNA, including detection of insertions/deletions as well as focal and broad copy number variations (CNVs). To address this unmet need, a targeted, hybrid capture-based next-generation sequencing platform covering a panel of 523 cancer-associated genes (TSO500ctDNATM), encompassing all major prognostic and driver mutations associated with pHGG, was employed.^{62,63} This approach successfully detected tumor-associated gene CNVs in DMG patient CSF, providing an opportunity for longitudinal gene CNV monitoring to understand the genomic drivers of pHGG evolution and disease progression.⁶¹

To further establish the clinical utility of ctDNA profiling, investigators in the ONC014 phase 1 trial of ONC201 in children and young adults (NCT03416530) conducted an arm with 24 patients with H3K27M-mutant DMG enrolled after radiation to undergo serial CSF collection.⁴⁵ Patients underwent serial lumbar puncture (LP) for cf-tDNA analysis at 0, 2, and 6 months on therapy, while patients enrolled at the University of Michigan underwent monthly serial plasma collection. ddPCR

analysis of cf-tDNA samples was performed, and variant allele fraction (VAF) was compared to radiographic measured by maximal 2D tumor area on magnetic resonance imaging (MRI). Patients were screened before LP to confirm clinical safety, with particular concern given to signs of increased intracranial pressure or rapidly changing neurologic exams. No adverse outcomes related to the procedure were observed.

Importantly, change in H3.3K27M VAF over time (“VAF delta”) correlated with prolonged PFS in both CSF and plasma samples, predicting progression and sustained response and possible differentiation of pseudo-progression and pseudo-response.⁶⁴ Indeed, VAF “spikes” (an increase of at least 25%) preceded tumor progression in many cases.⁶⁴

These results confirmed the feasibility and utility of serial cf-tDNA in both plasma and CSF of DMG patients to supplement radiographic monitoring. CSF allows for assessing other tumor biomarkers that may be enriched in spinal fluid, such as cell-free DNA methylation sequencing, exosomal RNA, and mitochondrial DNA (mtDNA). However, CSF collection carries more procedural risk than a blood draw, thus encouraging further research to optimize plasma diagnostics for DMG to complement or potentially replace CSF diagnostics. Multiple academic institutions and biotechnology companies are optimizing these methods and transitioning these tests to a CLIA-certified setting will allow for improved decision-making in the clinical management of DMG patients and potentially broader glioma patient populations.

Microbiome

The gut microbiome is vital in various diseases, including central nervous system diseases and cancer.^{65–68} For adult HGG, there is emerging evidence that the gut microbiome plays a role in pathogenesis.⁶⁹ Metabolites, such as tryptophan, are strongly influenced by the gut microbiome, and can modify glioma’s microenvironment by directly affecting T cells, dendritic cells, tumor-associated macrophages, and antigen-presenting cells.⁷⁰ Recent data on pediatric DMGs and the microbiome revealed a potential role of the microbiome on PFS and

OS. Specifically, the *Firmicutes/Bacteroidetes* ratio, which serves as a parameter of normal intestinal homeostasis, was observed to be unfavorable at diagnosis, and some components, such as Flavobacteriaceae and Bacillales, were associated with a higher risk of disease progression and death.⁷¹ To better understand and elucidate the underlying mechanisms of these observations, it is vital to gain insight into the microbial composition of trial patients with DMG, as is currently being performed in the PNOC22 clinical trials.

Predictive genomic biomarkers of ONC201 in DMG

DMGs harbor complex genomes with frequent somatic inactivation of *TP53*, chromosomal instability (e.g., loss of 10q), and high-level amplification of oncogenes (e.g., *PDGFRA*, *MET*, *EGFR*).⁷² We and others have demonstrated that ONC201 binds and activates ClpP, that *CLPP* depletion abrogates ONC201 sensitivity, and that *CLPP* expression levels predict ONC201 sensitivity in DMG and other cancer types.^{15,22,43,73} *CLPP* is located on 19p13.3, a region also susceptible to *TP53*-associated genomic instability in DMGs and thus potentially predictive of ONC201 sensitivity. Notably, protein-coding genes are dosage sensitive in cancer, and *CLPP* CNVs also explain *CLPP* expression differences between cancer cell lines (<https://depmap.org/portal/gene/CLPP>). Joint DMG whole genome and RNA sequencing in PNOC022 will inform whether high *CLPP* copy number and expression levels predict ONC201 sensitivity in DMG patients.

Studies in glioblastoma (GBM) and breast cancer provided evidence that ONC201 and ONC206 drive rapid depletion of mtDNA copy number, an effect not seen with other metabolic drugs such as metformin.^{74,75} Large-scale analysis of whole cancer genomes revealed an order of magnitude difference in mtDNA copy number between GBMs (~70 to 800 copies per cell).⁷⁶ So far, however, little is known to what extent mtDNA copy number varies between H3K27M-mutant and H3WT DMGs and whether it will modulate ONC201 sensitivity. We will address this question in PNOC022 based on quantification of mtDNA copy number levels from DMG whole genomes using pipelines developed within the Pan-Cancer Analysis of Whole Genomes Consortium.⁷⁶

Oncogenic signaling pathways have also been nominated to confer resistance to ONC201. In 2021, He and colleagues observed that high expression levels of *EGFR*, the constitutively active form of EGFR (EGFRvIII), and EGFR staining in GBM are determinants of poor pre-clinical and clinical response to ONC201.⁷⁷ A recent independent effort to identify combinatorial biomarkers using transcriptomics and proteomics in a large panel of cancer cell lines has also highlighted ClpP and EGFR as predictors of ONC201 and ONC206 sensitivity.⁷⁸ A similar ONC201 resistance mechanism is also studied in H3K27M-mutant DMGs that harbor oncogenic *EGFR* mutations, high-level *EGFR* amplifications, or high *EGFR* expression levels and will be addressed based on RNA- and whole genome sequencing in PNOC022.⁷⁹

H3K27M-mutant DMGs are likely stalled in a cellular state between neural stem cells and oligodendrocyte precursor cells (OPC) with a subpopulation of DMGs being defined by high levels of *EGFR* expression.⁵⁶⁻⁵⁸ This subpopulation resembles a developmental *EGFR*-high intermediate progenitor cell state termed Pre-OPC⁸⁰⁻⁸³ and we observed that ONC201 drives the differentiation of OPC-like DMG cells towards other cell states which raises the possibility of a non-genetic and cell-intrinsic ONC201-resistant subpopulation of Pre-OPC-like DMG cells, thus necessitating combination therapies such as ONC201 and paxalisib.¹⁵

Quality of life and patient-reported outcomes

Along with toxicity, efficacy, and biomarker assessments, PNOC022 aims to collect details on patient experience throughout participation in the trial. Patient-reported outcomes (PROs) are reports of the status of a patient's health condition that comes directly from the patient and/or proxy (in patients less than 18 years of age) without interpretation of the response by a clinician.⁸⁶ PROs are increasingly acknowledged as an essential source of information about patient and/or proxy experience during cancer treatment. These assessments allow patients/proxies to report on side effects and overall quality-of-life during and after treatment; as such, they are being utilized as direct measures of outcomes in clinical trials.⁸⁷⁻⁹⁵ PROs may provide insight into how patients/proxies perceive the care they have received, and can be administered longitudinally throughout the clinical course. Consensus as to which measures to include within clinical trials is beginning to come into focus for various patient populations. For example, a recent publication from the National Clinical Trials Network (NCTN) Adolescent and Young Adult (AYA) PRO Task Force put forth a

recommended core battery of PRO measures to include in clinical trials targeting the AYA population. This battery has inherent flexibility, allowing the PROs to shift based on the trial population and specific study objectives. There are few prospective measures of patient/proxy perception of and satisfaction with participation in a clinical trial, with almost no prospective data within the pediatric cancer patient population.⁹³⁻⁹⁵ Within PNOC022, we hypothesized that there will be at least some perceived benefit from participation in PNOC022 and that the perceived benefit, or lack thereof, of study participation will be variable over time, depending on the intensity of the phase of therapy at the time of questionnaire administration.

To understand how the patient and/or proxy perceives trial participation, we created a four-question PRO (“DMG-ACT PRO”). The DMG-ACT PRO is administered at post-resection/biopsy; end of radiation; beginning of cycles 3, 6, 12, 18, and 24; and end of treatment. The questions offer insight into the overall satisfaction of study participation, the likelihood of recommending DMG-ACT or a similar study to a future patient, and the retrospective likelihood of participating again in this study. The final question centers around why the patient/proxy answered as they did about the retrospective likelihood of participating in the study with four pre-written choices and an option for free text response.

Consideration of novel imaging assessment integration

Standard contrast-enhanced MRI is the primary clinical imaging modality to determine treatment response.⁹⁶ However, the conventional MRI technique has its limitations. For example, differentiation between pseudo-progression or tumor progression/recurrence can be difficult. In DMGs, pseudo-progression is commonly seen and has been described in up to 50% of patients.^{97,98} This can cause a clinical dilemma with the risk of early termination of treatment in a clinical trial. Therefore, further optimization of imaging techniques is needed and advanced MRI techniques and positron emission tomography (PET) may be helpful.

Advanced MRI techniques

Advanced MRI methods that are now available within standard MR sequencing options, including MR perfusion (MRP) and MR spectroscopy (MRS), have emerged. MRP is used for imaging the vascularity of the brain and the tumor, with different parameters such as arterial-spin labeling (ASL) and cerebral blood flow (CBF). In DMG, the prognostic value of MRP has been demonstrated.⁹⁹ For instance, increased ASL-CBF can be observed in pseudo-progression compared to actual progression.⁹⁷ Further, a difference in perfusion values between tumors with H3.1K27M and H3.3K27M mutations were found in pediatric DMGs, suggesting that this imaging modality may offer prognostic and therapeutic insights.⁹⁷ MRS measures biochemical changes in the brain, and the most commonly used technique is proton (¹H; hydrogen) spectroscopy. Cellular metabolites are measured, such as choline (Cho), creatine, lactate, lipids, and myoinositol. The biochemical profiles vary by brain regions and with brain maturation. Proton MRS can help differentiate types of brain tumors, especially those found in the infratentorial compartment, by looking at ratios of N-acetyl aspartate (NAA), creatine, Cho, and lactate.¹⁰⁰

In one prospective study of 36 patients with pontine DMGs after radiation therapy, it was found that patients with higher Cho: NAA values were at greater mortality risk. Additionally, an increase in Cho: NAA inversely correlated with survival throughout the disease course.¹⁰¹ Further efforts are needed to determine what metabolic measures on ¹H-MRS at diagnosis are the best survival or treatment response predictors.¹⁰⁰ Indeed, although the diagnostic value of MRS in differentiating between tumor recurrence or radiation necrosis has been shown in a systematic review of 28 studies covering adult brain tumors, similar work is currently absent for DMGs.¹⁰²

PET/¹⁸F-FDG

PET visualizes metabolic features, which can assist in determining the most active tumor location, thereby guiding a biopsy's target tissue and discriminating between pseudo-progression and tumor progression. Ideally, PET and MRI scans are performed in a single session on a hybrid PET/MRI system in pediatric patients, minimizing the diagnostic burden for the patient.

^{18}F -fluorodeoxyglucose (^{18}F -FDG) was one of the first radiotracer used for PET imaging. Unfortunately, however, the normal brain has a physiologic intense ^{18}F -FDG uptake, which limits the sensitivity for detecting brain lesions. This contributes to limited ^{18}F -FDG activity in DMG and, subsequently, contradictory results regarding the correlation between PET parameters and survival in this disease.⁹⁹

In recent years, radiolabeled amino acids such as ^{11}C -methyl-L-methionine (^{11}C -MET), ^{18}F -fluoroethyl-L-tyrosine (^{18}F -FET), and ^{18}F -fluoro-L-dihydroxyphenylalanine (^{18}F -DOPA) have been widely introduced for brain tumor imaging and are some of the most commonly applied radiotracers.¹⁰³ Importantly, these tracers have a minimal uptake in normal brain compared to ^{18}F -FDG, each capable of crossing the BBB. Their uptake depends on the expression of the amino acid transporter, L-type amino acid transporter 1.⁹⁹

^{11}C -MET is the oldest amino acid tracer used in brain tumor imaging. Few studies have been performed in DMG patients but have shown that ^{11}C -MET-PET can visualize the tumors in most cases.^{104,105} Although ^{11}C -MET-PET has proven its clinical value, ^{18}F -FET-PET has logistic advantages over ^{11}C -MET-PET because of its longer half-life, allowing for dynamic scanning with the evaluation of time-activity-curves (TACs) and independence from a cyclotron.

^{18}F -DOPA has been studied in children with DMGs, with ^{18}F -DOPA-PET capable of differentiating between H3K27M-mutant and H3WT DMGs and predicting outcomes.^{106,107} A recent retrospective study in 15 pediatric brain tumor patients, of which five had confirmed H3K27-altered DMG and 2 had a clinical/radiologic diagnosis of pontine DMG, analyzed the added value of dynamic ^{18}F -DOPA-PET and computed tomography parameters.¹⁰⁸ In this study, the TAC dynamic PET parameter identified patients at higher risk of disease progression and death.

In adults with gliomas, ^{18}F -FET-PET is widely used, and evidence-based guidelines are described by the EANM/EANO/RANO working group.¹⁰⁹ In 2015, Dunkl et al. described the first cohort of pediatric patients with brain tumors who received dynamic ^{18}F -FET-PET for further diagnostic scanning because of complex initial imaging findings on MRI.¹¹⁰ They found that this

approach may be helpful for the identification of newly diagnosed brain lesions suggestive of glioma and in the diagnosis of tumor progression or recurrence.

A case report from an adult with an H3K27M-mutant DMG showed high ^{18}F -FET uptake and a short time to peak.¹¹¹ Specific to the pediatric population, the diagnostic accuracy and clinical impact of ^{18}F -FET have been reported in a prospective study with 169 scans in 97 children and adolescents comprising a variety of high- and low-grade tumors, including six children with H3K27M-mutant DMG.¹¹² Indeed, ^{18}F -FET-PET showed significantly higher accuracy for detecting tumors in both untreated and treated lesions than MRI and altered the treatment plan in 33% of patients with a clinical indication for additional imaging.

Advanced MRI and PET imaging can offer additional value in managing DMG, including use at diagnosis to define the best target for biopsy and assistance in radiation planning. During treatment, it may correlate with survival or treatment response and discriminate between treatment-related effects versus tumor progression. However, a larger, homogenous cohort of DMG patients needs to be studied with sequential scan time points to prove further the additional value of metabolic imaging approaches, such as ^{18}F -FET-PET, and determine how these can be integrated into clinical care. By incorporating ^{18}F -FET-PET imaging into PNOC022, we will be able to address some of these questions and also assess utility within a clinical trial setting for children with DMGs.

Conclusions

DMG remains a deadly disease that deserves thoughtful, multidisciplinary, and multifaceted clinical trial design in the quest to find a cure. PNOC022 provides an adaptive platform that allows near real-time incorporation of new treatment arms based on evolving understanding of potential disease vulnerabilities. The trial also incorporates collection of novel pharmacokinetic, molecular, genomic, and imaging biomarkers that will inform on tumors' mechanisms of response and resistance. Each piece of the trial has been developed with expertise from basic scientists to clinical researchers to pharmacologists to imaging specialists. Further, the PNOC mission includes trial access for as many

children and young adults as feasible, regardless of geographic location. To that end, PNOC022 is currently open at 31 sites across five countries, including several sites in Australia/New Zealand and Israel. Through the design and collaboration of PNOC022, we will greatly augment our understanding of DMG with the ultimate goal to advance more effective therapy combinations in rapid pace.

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References

1. Jansen MH, Veldhuijzen van Zanten SE, Sanchez Aliaga E, et al. Survival prediction model of children with diffuse intrinsic pontine glioma based on clinical and radiological criteria. *Neuro-Oncol.* 2015;17(1):160-166. doi:10.1093/neuonc/nou104
2. Findlay IJ, De Iuliis GN, Duchatel RJ, et al. Pharmaco-proteogenomic profiling of pediatric diffuse midline glioma to inform future treatment strategies. *Oncogene.* 2022;41(4):461-475. doi:10.1038/s41388-021-02102-y
3. Warren KE. Diffuse intrinsic pontine glioma: poised for progress. *Front Oncol.* 2012;2. doi:10.3389/fonc.2012.00205
4. Schwartzenruber J, Korshunov A, Liu XY, et al. Driver mutations in histone H3.3 and chromatin remodelling genes in paediatric glioblastoma. *Nature.* 2012;482(7384):226-231. doi:10.1038/nature10833
5. St. Jude Children's Research Hospital–Washington University Pediatric Cancer Genome Project. Somatic histone H3 alterations in pediatric diffuse intrinsic pontine gliomas and non-brainstem glioblastomas. *Nat Genet.* 2012;44(3):251-253. doi:10.1038/ng.1102
6. Buczkowicz P, Hoeman C, Rakopoulos P, et al. Genomic analysis of diffuse intrinsic pontine gliomas identifies three molecular subgroups and recurrent activating ACVR1 mutations. *Nat Genet.* 2014;46(5):451-456. doi:10.1038/ng.2936
7. Khuong-Quang DA, Buczkowicz P, Rakopoulos P, et al. K27M mutation in histone H3.3 defines clinically and biologically distinct subgroups of pediatric diffuse intrinsic pontine gliomas. *Acta Neuropathol (Berl).* 2012;124(3):439-447. doi:10.1007/s00401-012-0998-0
8. Bender S, Tang Y, Lindroth AM, et al. Reduced H3K27me3 and DNA Hypomethylation Are Major Drivers of Gene Expression in K27M Mutant Pediatric High-Grade Gliomas. *Cancer Cell.* 2013;24(5):660-672. doi:10.1016/j.ccr.2013.10.006
9. Pasini D, Malatesta M, Jung HR, et al. Characterization of an antagonistic switch between histone H3 lysine 27 methylation and acetylation in the transcriptional regulation of Polycomb group target genes. *Nucleic Acids Res.* 2010;38(15):4958-4969. doi:10.1093/nar/gkq244
10. Piunti A, Hashizume R, Morgan MA, et al. Therapeutic targeting of polycomb and BET bromodomain proteins in diffuse intrinsic pontine gliomas. *Nat Med.* 2017;23(4):493-500. doi:10.1038/nm.4296
11. Larson JD, Kasper LH, Paugh BS, et al. Histone H3.3 K27M Accelerates Spontaneous Brainstem Glioma and Drives Restricted Changes in Bivalent Gene Expression. *Cancer Cell.* 2019;35(1):140-155.e7. doi:10.1016/j.ccell.2018.11.015
12. Jain SU, Rashoff AQ, Krabbenhoft SD, et al. H3 K27M and EZHIP Impede H3K27-Methylation Spreading by Inhibiting Allosterically Stimulated PRC2. *Mol Cell.* 2020;80(4):726-735.e7. doi:10.1016/j.molcel.2020.09.028

13. Krug B, De Jay N, Harutyunyan AS, et al. Pervasive H3K27 Acetylation Leads to ERV Expression and a Therapeutic Vulnerability in H3K27M Gliomas. *Cancer Cell*. 2019;35(5):782-797.e8. doi:10.1016/j.ccell.2019.04.004
14. Golbourn BJ, Halbert ME, Halligan K, et al. Loss of MAT2A compromises methionine metabolism and represents a vulnerability in H3K27M mutant glioma by modulating the epigenome. *Nat Cancer*. 2022;3(5):629-648. doi:10.1038/s43018-022-00348-3
15. Przystal JM, Cianciolo Cosentino C, Yadavilli S, et al. Imipridones affect tumor bioenergetics and promote cell lineage differentiation in diffuse midline gliomas. *Neuro-Oncol*. Published online February 14, 2022: noac041. doi:10.1093/neuonc/noac041
16. Cole A, Wang Z, Coyaud E, et al. Inhibition of the Mitochondrial Protease ClpP as a Therapeutic Strategy for Human Acute Myeloid Leukemia. *Cancer Cell*. 2015;27(6):864-876. doi:10.1016/j.ccell.2015.05.004
17. Conlon BP, Nakayasu ES, Fleck LE, et al. Activated ClpP kills persisters and eradicates a chronic biofilm infection. *Nature*. 2013;503(7476):365-370. doi:10.1038/nature12790
18. De Sagarra MR, Mayo I, Marco S, et al. Mitochondrial localization and oligomeric structure of HClpP, the human homologue of E. coli ClpP. *J Mol Biol*. 1999;292(4):819-825. doi:10.1006/jmbi.1999.3121
19. Graves PR, Aponte-Collazo LJ, Fennell EMJ, et al. Mitochondrial Protease ClpP is a Target for the Anticancer Compounds ONC201 and Related Analogues. *ACS Chem Biol*. 2019;14(5):1020-1029. doi:10.1021/acscchembio.9b00222
20. Ishizawa J, Zarabi SF, Davis RE, et al. Mitochondrial ClpP-Mediated Proteolysis Induces Selective Cancer Cell Lethality. *Cancer Cell*. 2019;35(5):721-737.e9. doi:10.1016/j.ccell.2019.03.014
21. Jacques S, Van Der Sloot AM, C. Huard C, et al. Imipridone Anticancer Compounds Ectopically Activate the ClpP Protease and Represent a New Scaffold for Antibiotic Development. *Genetics*. 2020;214(4):1103-1120. doi:10.1534/genetics.119.302851
22. Bonner ER, Waszak SM, Grotzer MA, Mueller S, Nazarian J. Mechanisms of imipridones in targeting mitochondrial metabolism in cancer cells. *Neuro-Oncol*. 2021;23(4):542-556. doi:10.1093/neuonc/noaa283
23. Chari A, Barlogie B. Imipridone ONC201: combination therapy in hematologic malignancies. *Cell Cycle*. 2018;17(16):1947-1948. doi:10.1080/15384101.2018.1490861
24. Edwards H, Ge Y. ONC201 shows promise in AML treatment. *Cell Cycle*. 2018;17(3):277-277. doi:10.1080/15384101.2017.1421035
25. Lev A, Lulla AR, Ross BC, et al. ONC201 Targets AR and AR-V7 Signaling, Reduces PSA, and Synergizes with Everolimus in Prostate Cancer. *Mol Cancer Res*. 2018;16(5):754-766. doi:10.1158/1541-7786.MCR-17-0614
26. Arrillaga-Romany I, Odia Y, Prabhu VV, et al. Biological activity of weekly ONC201 in adult recurrent glioblastoma patients. *Neuro-Oncol*. 2020;22(1):94-102. doi:10.1093/neuonc/noz164

27. Prabhu VV, Morrow S, Rahman Kawakibi A, et al. ONC201 and imipridones: Anti-cancer compounds with clinical efficacy. *Neoplasia*. 2020;22(12):725-744. doi:10.1016/j.neo.2020.09.005
28. Venneti S, Kawakibi AR, Ji S, et al. Clinical efficacy of ONC201 in H3K27M-mutant diffuse midline gliomas is driven by disruption of integrated metabolic and epigenetic pathways. *Cancer Discov*. Published online August 16, 2023:CD-23-0131. doi:10.1158/2159-8290.CD-23-0131
29. Chung C, Sweha SR, Pratt D, et al. Integrated Metabolic and Epigenomic Reprogramming by H3K27M Mutations in Diffuse Intrinsic Pontine Gliomas. *Cancer Cell*. 2020;38(3):334-349.e9. doi:10.1016/j.ccell.2020.07.008
30. Magaway C, Kim E, Jacinto E. Targeting mTOR and Metabolism in Cancer: Lessons and Innovations. *Cells*. 2019;8(12):1584. doi:10.3390/cells8121584
31. Georgescu MM, Islam MZ, Li Y, et al. Global activation of oncogenic pathways underlies therapy resistance in diffuse midline glioma. *Acta Neuropathol Commun*. 2020;8(1):111. doi:10.1186/s40478-020-00992-9
32. Paugh BS, Broniscer A, Qu C, et al. Genome-Wide Analyses Identify Recurrent Amplifications of Receptor Tyrosine Kinases and Cell-Cycle Regulatory Genes in Diffuse Intrinsic Pontine Glioma. *J Clin Oncol*. 2011;29(30):3999-4006. doi:10.1200/JCO.2011.35.5677
33. Makinoshima H, Takita M, Saruwatari K, et al. Signaling through the Phosphatidylinositol 3-Kinase (PI3K)/Mammalian Target of Rapamycin (mTOR) Axis Is Responsible for Aerobic Glycolysis mediated by Glucose Transporter in Epidermal Growth Factor Receptor (EGFR)-mutated Lung Adenocarcinoma. *J Biol Chem*. 2015;290(28):17495-17504. doi:10.1074/jbc.M115.660498
34. Ribback S, Cigliano A, Kroeger N, et al. PI3K/AKT/mTOR pathway plays a major pathogenetic role in glycogen accumulation and tumor development in renal distal tubules of rats and men. *Oncotarget*. 2015;6(15):13036-13048. doi:10.18632/oncotarget.3675
35. Wang X, Lin Y, Kemper T, et al. AMPK and Akt/mTOR signalling pathways participate in glucose-mediated regulation of hepatitis B virus replication and cellular autophagy. *Cell Microbiol*. 2020;22(2). doi:10.1111/cmi.13131
36. Aldonza MBD, Hong JY, Bae SY, et al. Suppression of MAPK Signaling and Reversal of mTOR-Dependent MDR1-Associated Multidrug Resistance by 21 α -Methylmelanodiol in Lung Cancer Cells. Castresana JS, ed. *PLOS ONE*. 2015;10(6):e0127841. doi:10.1371/journal.pone.0127841
37. Poore B, Yuan M, Arnold A, et al. Inhibition of mTORC1 in pediatric low-grade glioma depletes glutathione and therapeutically synergizes with carboplatin. *Neuro-Oncol*. 2019;21(2):252-263. doi:10.1093/neuonc/noy150
38. Montes DK, Brenet M, Muñoz VC, et al. Vasopressin activates Akt/mTOR pathway in smooth muscle cells cultured in high glucose concentration. *Biochem Biophys Res Commun*. 2013;441(4):923-928. doi:10.1016/j.bbrc.2013.10.169

39. Heffron TP, Ndubaku CO, Salphati L, et al. Discovery of Clinical Development Candidate GDC-0084, a Brain Penetrant Inhibitor of PI3K and mTOR. *ACS Med Chem Lett.* 2016;7(4):351-356. doi:10.1021/acsmchemlett.6b00005
40. He C, Xu K, Zhu X, et al. Patient-derived models recapitulate heterogeneity of molecular signatures and drug response in pediatric high-grade glioma. *Nat Commun.* 2021;12(1):4089. doi:10.1038/s41467-021-24168-8
41. Wen PY, de Groot JF, Battiste J, et al. Paxalisib in patients with newly diagnosed glioblastoma with unmethylated MGMT promoter status: Final phase 2 study results. *J Clin Oncol.* 2022;40(16_suppl):2047-2047. doi:10.1200/JCO.2022.40.16_suppl.2047
42. Wen PY, de Groot J, Battiste JD, et al. Abstract LB125: Pharmacokinetics of paxalisib in phase 2 clinical study in glioblastoma (GBM) with unmethylated O6-methylguanine-methyltransferase (MGMT) promoter status. *Cancer Res.* 2021;81(13_Supplement):LB125-LB125. doi:10.1158/1538-7445.AM2021-LB125
43. Jackson ER, Duchatel RJ, Staudt DE, et al. ONC201 in combination with paxalisib for the treatment of H3K27-altered diffuse midline glioma. *Cancer Res.* Published online May 5, 2023:CAN-23-0186. doi:10.1158/0008-5472.CAN-23-0186
44. Tang R, Shen J, Yuan Y. ComPAS: A Bayesian drug combination platform trial design with adaptive shrinkage. *Stat Med.* 2019;38(7):1120-1134. doi:10.1002/sim.8026
45. Gardner SL, Allen JC, Zaky WT, et al. ONC201 in previously-irradiated pediatric H3 K27M-mutant glioma. *J Clin Oncol.* 2019;37(15_suppl):10046-10046. doi:10.1200/JCO.2019.37.15_suppl.10046
46. Lassaletta A, Strother D, Laperriere N, et al. Reirradiation in patients with diffuse intrinsic pontine gliomas: The Canadian experience. *Pediatr Blood Cancer.* 2018;65(6):e26988. doi:10.1002/pbc.26988
47. Morales La Madrid A, Santa-María V, Cruz Martinez O, et al. Second re-irradiation for DIPG progression, re-considering “old strategies” with new approaches. *Childs Nerv Syst.* 2017;33(5):849-852. doi:10.1007/s00381-017-3352-y
48. Tsang DS, Oliveira C, Bouffet E, et al. Repeat irradiation for children with supratentorial high- grade glioma. *Pediatr Blood Cancer.* 2019;66(9). doi:10.1002/pbc.27881
49. Kline C, Liu SJ, Duriseti S, et al. Reirradiation and PD-1 inhibition with nivolumab for the treatment of recurrent diffuse intrinsic pontine glioma: a single-institution experience. *J Neurooncol.* 2018;140(3):629-638. doi:10.1007/s11060-018-2991-5
50. Müller K, Scheithauer H, Pietschmann S, et al. Reirradiation as part of a salvage treatment approach for progressive non-pontine pediatric high-grade gliomas: preliminary experiences from the German HIT-HGG study group. *Radiat Oncol.* 2014;9(1):177. doi:10.1186/1748-717X-9-177
51. Freese C, Takiar V, Fouladi M, DeWire M, Breneman J, Pater L. Radiation and subsequent reirradiation outcomes in the treatment of diffuse intrinsic pontine glioma and a systematic review of the reirradiation literature. *Pract Radiat Oncol.* 2017;7(2):86-92. doi:10.1016/j.pro.2016.11.005

52. Wang Y, Springer S, Zhang M, et al. Detection of tumor-derived DNA in cerebrospinal fluid of patients with primary tumors of the brain and spinal cord. *Proc Natl Acad Sci.* 2015;112(31):9704-9709. doi:10.1073/pnas.1511694112
53. Bruzek AK, Ravi K, Muruganand A, et al. Electronic DNA Analysis of CSF Cell-free Tumor DNA to Quantify Multi-gene Molecular Response in Pediatric High-grade Glioma. *Clin Cancer Res.* 2020;26(23):6266-6276. doi:10.1158/1078-0432.CCR-20-2066
54. Miller AM, Shah RH, Pentsova EI, et al. Tracking tumour evolution in glioma through liquid biopsies of cerebrospinal fluid. *Nature.* 2019;565(7741):654-658. doi:10.1038/s41586-019-0882-3
55. Liu APY, Smith KS, Kumar R, et al. Serial assessment of measurable residual disease in medulloblastoma liquid biopsies. *Cancer Cell.* 2021;39(11):1519-1530.e4. doi:10.1016/j.ccell.2021.09.012
56. Huang TY, Piunti A, Lulla RR, et al. Detection of Histone H3 mutations in cerebrospinal fluid-derived tumor DNA from children with diffuse midline glioma. *Acta Neuropathol Commun.* 2017;5(1):28. doi:10.1186/s40478-017-0436-6
57. Izquierdo E, Proszek P, Pericoli G, et al. Droplet digital PCR-based detection of circulating tumor DNA from pediatric high grade and diffuse midline glioma patients. *Neuro-Oncol Adv.* 2021;3(1):vdab013. doi:10.1093/oaajnl/vdab013
58. Stallard S, Savelieff MG, Wierzbicki K, et al. CSF H3F3A K27M circulating tumor DNA copy number quantifies tumor growth and in vitro treatment response. *Acta Neuropathol Commun.* 2018;6(1):80. doi:10.1186/s40478-018-0580-7
59. Panditharatna E, Kilburn LB, Aboian MS, et al. Clinically Relevant and Minimally Invasive Tumor Surveillance of Pediatric Diffuse Midline Gliomas Using Patient-Derived Liquid Biopsy. *Clin Cancer Res.* 2018;24(23):5850-5859. doi:10.1158/1078-0432.CCR-18-1345
60. Li D, Bonner ER, Wierzbicki K, et al. Standardization of the liquid biopsy for pediatric diffuse midline glioma using ddPCR. *Sci Rep.* 2021;11(1):5098. doi:10.1038/s41598-021-84513-1
61. Bonner ER, Harrington R, Eze A, et al. Circulating tumor DNA sequencing provides comprehensive mutation profiling for pediatric central nervous system tumors. *Npj Precis Oncol.* 2022;6(1):63. doi:10.1038/s41698-022-00306-3
62. Mueller S, Jain P, Liang WS, et al. A pilot precision medicine trial for children with diffuse intrinsic pontine glioma—PNOC003: A report from the Pacific Pediatric Neuro-Oncology Consortium. *Int J Cancer.* Published online April 3, 2019:ijc.32258. doi:10.1002/ijc.32258
63. Nikbakht H, Panditharatna E, Mikael LG, et al. Spatial and temporal homogeneity of driver mutations in diffuse intrinsic pontine glioma. *Nat Commun.* 2016;7(1):11185. doi:10.1038/ncomms11185

64. Cantor E, Wierzbicki K, Tarapore RS, et al. Serial H3K27M cell-free tumor DNA (cf-tDNA) tracking predicts ONC201 treatment response and progression in diffuse midline glioma. *Neuro-Oncol.* 2022;24(8):1366-1374. doi:10.1093/neuonc/noac030
65. Sepich-Poore GD, Zitvogel L, Straussman R, Hasty J, Wargo JA, Knight R. The microbiome and human cancer. *Science.* 2021;371(6536):eabc4552. doi:10.1126/science.abc4552
66. Li J, Zhao S, Lee M, et al. Reliable tumor detection by whole-genome methylation sequencing of cell-free DNA in cerebrospinal fluid of pediatric medulloblastoma. *Sci Adv.* 2020;6(42):eabb5427. doi:10.1126/sciadv.abb5427
67. Welton JL, Loveless S, Stone T, von Ruhland C, Robertson NP, Clayton A. Cerebrospinal fluid extracellular vesicle enrichment for protein biomarker discovery in neurological disease; multiple sclerosis. *J Extracell Vesicles.* 2017;6(1):1369805. doi:10.1080/20013078.2017.1369805
68. Peng Y, Zheng D, Zhang X, et al. Cell-Free Mitochondrial DNA in the CSF: A Potential Prognostic Biomarker of Anti-NMDAR Encephalitis. *Front Immunol.* 2019;10:103. doi:10.3389/fimmu.2019.00103
69. Nejman D, Livyatan I, Fuks G, et al. The human tumor microbiome is composed of tumor type-specific intracellular bacteria. *Science.* 2020;368(6494):973-980. doi:10.1126/science.aay9189
70. Liang J, Li T, Zhao J, Wang C, Sun H. Current understanding of the human microbiome in glioma. *Front Oncol.* 2022;12:781741. doi:10.3389/fonc.2022.781741
71. De Cecco L, Biassoni V, Schiavello E, et al. DIPG-36. The brain-gut-microbiota axis to predict outcome in pediatric diffuse intrinsic pontine glioma. *Neuro-Oncol.* 2022;24(Supplement_1):i26-i26. doi:10.1093/neuonc/noac079.093
72. Kline C, Jain P, Kilburn L, et al. Upfront Biology-Guided Therapy in Diffuse Intrinsic Pontine Glioma: Therapeutic, Molecular, and Biomarker Outcomes from PNOC003. *Clin Cancer Res.* 2022;28(18):3965-3978. doi:10.1158/1078-0432.CCR-22-0803
73. Fennell EMJ, Aponte-Collazo LJ, Pathmasiri W, et al. Multi-omics analyses reveal ClpP activators disrupt essential mitochondrial pathways in triple-negative breast cancer. *Front Pharmacol.* 2023;14:1136317. doi:10.3389/fphar.2023.1136317
74. Ishida CT, Zhang Y, Bianchetti E, et al. Metabolic Reprogramming by Dual AKT/ERK Inhibition through Imipridones Elicits Unique Vulnerabilities in Glioblastoma. *Clin Cancer Res Off J Am Assoc Cancer Res.* 2018;24(21):5392-5406. doi:10.1158/1078-0432.CCR-18-1040
75. Greer YE, Porat-Shliom N, Nagashima K, et al. ONC201 kills breast cancer cells in vitro by targeting mitochondria. *Oncotarget.* 2018;9(26):18454-18479. doi:10.18632/oncotarget.24862
76. Yuan Y, Ju YS, Kim Y, et al. Comprehensive molecular characterization of mitochondrial genomes in human cancers. *Nat Genet.* 2020;52(3):342-352. doi:10.1038/s41588-019-0557-x

77. He Y, Li J, Koga T, et al. Epidermal growth factor receptor as a molecular determinant of glioblastoma response to dopamine receptor D2 inhibitors. *Neuro-Oncol.* 2021;23(3):400-411. doi:10.1093/neuonc/noaa188
78. Morrow S, Nath K, Zhang Y, et al. Abstract 393: Predictive biomarker evaluation and molecular differentiation for imipridones ONC201 and ONC206. *Cancer Res.* 2021;81(13_Supplement):393-393. doi:10.1158/1538-7445.AM2021-393
79. Kawakibi AR, Tarapore R, Gardner S, et al. CTNI-61. CLINICAL EFFICACY AND PREDICTIVE BIOMARKERS OF ONC201 IN H3K27M-MUTANT DIFFUSE MIDLINE GLIOMA. *Neuro-Oncol.* 2022;24(Supplement_7):vii86-vii87. doi:10.1093/neuonc/noac209.326
80. Liu I, Jiang L, Samuelsson ER, et al. The landscape of tumor cell states and spatial organization in H3-K27M mutant diffuse midline glioma across age and location. *Nat Genet.* 2022;54(12):1881-1894. doi:10.1038/s41588-022-01236-3
81. Huang W, Bhaduri A, Velmeshev D, et al. Origins and Proliferative States of Human Oligodendrocyte Precursor Cells. *Cell.* 2020;182(3):594-608.e11. doi:10.1016/j.cell.2020.06.027
82. Trevino AE, Müller F, Andersen J, et al. Chromatin and gene-regulatory dynamics of the developing human cerebral cortex at single-cell resolution. *Cell.* 2021;184(19):5053-5069.e23. doi:10.1016/j.cell.2021.07.039
83. Andersen J, Thom N, Shadrach JL, et al. Single-cell transcriptomic landscape of the developing human spinal cord. *Nat Neurosci.* 2023;26(5):902-914. doi:10.1038/s41593-023-01311-w
84. Filbin MG, Tirosh I, Hovestadt V, et al. Developmental and oncogenic programs in H3K27M gliomas dissected by single-cell RNA-seq. *Science.* 2018;360(6386):331-335. doi:10.1126/science.aao4750
85. Jessa S, Blanchet-Cohen A, Krug B, et al. Stalled developmental programs at the root of pediatric brain tumors. *Nat Genet.* 2019;51(12):1702-1713. doi:10.1038/s41588-019-0531-7
86. Thorlund K, Dron L, Park JJ, Mills EJ. Synthetic and External Controls in Clinical Trials – A Primer for Researchers. *Clin Epidemiol.* 2020;Volume 12:457-467. doi:10.2147/CLEP.S242097
87. Coens C, Pe M, Dueck AC, et al. International standards for the analysis of quality-of-life and patient-reported outcome endpoints in cancer randomised controlled trials: recommendations of the SISAQOL Consortium. *Lancet Oncol.* 2020;21(2):e83-e96. doi:10.1016/S1470-2045(19)30790-9
88. Basch E. Missing Patients' Symptoms in Cancer Care Delivery--The Importance of Patient-Reported Outcomes. *JAMA Oncol.* 2016;2(4):433-434. doi:10.1001/jamaoncol.2015.4719

89. Leahy AB, Steineck A. Patient-Reported Outcomes in Pediatric Oncology: The Patient Voice as a Gold Standard. *JAMA Pediatr.* 2020;174(11):e202868. doi:10.1001/jamapediatrics.2020.2868
90. Reeve BB, Withycombe JS, Baker JN, et al. The first step to integrating the child's voice in adverse event reporting in oncology trials: a content validation study among pediatric oncology clinicians. *Pediatr Blood Cancer.* 2013;60(7):1231-1236. doi:10.1002/pbc.24463
91. Leahy AB, Feudtner C, Basch E. Symptom Monitoring in Pediatric Oncology Using Patient-Reported Outcomes: Why, How, and Where Next. *The Patient.* 2018;11(2):147-153. doi:10.1007/s40271-017-0279-z
92. Dobrozsi S, Yan K, Hoffmann R, Panepinto J. Patient-reported health status during pediatric cancer treatment. *Pediatr Blood Cancer.* 2017;64(4). doi:10.1002/pbc.26295
93. Roth ME, Parsons SK, Ganz PA, et al. Inclusion of a core patient-reported outcomes battery in adolescent and young adult cancer clinical trials. *J Natl Cancer Inst.* 2023;115(1):21-28. doi:10.1093/jnci/djac166
94. Cheng JD, Hitt J, Koczwara B, et al. Impact of quality of life on patient expectations regarding phase I clinical trials. *J Clin Oncol Off J Am Soc Clin Oncol.* 2000;18(2):421-428. doi:10.1200/JCO.2000.18.2.421
95. DasMahapatra P, Raja P, Gilbert J, Wicks P. Clinical trials from the patient perspective: survey in an online patient community. *BMC Health Serv Res.* 2017;17(1):166. doi:10.1186/s12913-017-2090-x
96. Cooney TM, Cohen KJ, Guimaraes CV, et al. Response assessment in diffuse intrinsic pontine glioma: recommendations from the Response Assessment in Pediatric Neuro-Oncology (RAPNO) working group. *Lancet Oncol.* 2020;21(6):e330-e336. doi:10.1016/S1470-2045(20)30166-2
97. Calmon R, Puget S, Varlet P, et al. Cerebral blood flow changes after radiation therapy identifies pseudoprogression in diffuse intrinsic pontine gliomas. *Neuro-Oncol.* 2018;20(7):994-1002. doi:10.1093/neuonc/nox227
98. Carceller F, Fowkes LA, Khabra K, et al. Pseudoprogression in children, adolescents and young adults with non-brainstem high grade glioma and diffuse intrinsic pontine glioma. *J Neurooncol.* 2016;129(1):109-121. doi:10.1007/s11060-016-2151-8
99. Lovibond S, Gewirtz AN, Pasquini L, Krebs S, Graham MS. The promise of metabolic imaging in diffuse midline glioma. *Neoplasia.* 2023;39:100896. doi:10.1016/j.neo.2023.100896
100. Blüml S, Saunders A, Tamrazi B. Proton MR Spectroscopy of Pediatric Brain Disorders. *Diagnostics.* 2022;12(6):1462. doi:10.3390/diagnostics12061462
101. Steffen-Smith EA, Shih JH, Hipp SJ, Bent R, Warren KE. Proton magnetic resonance spectroscopy predicts survival in children with diffuse intrinsic pontine glioma. *J Neurooncol.* 2011;105(2):365-373. doi:10.1007/s11060-011-0601-x

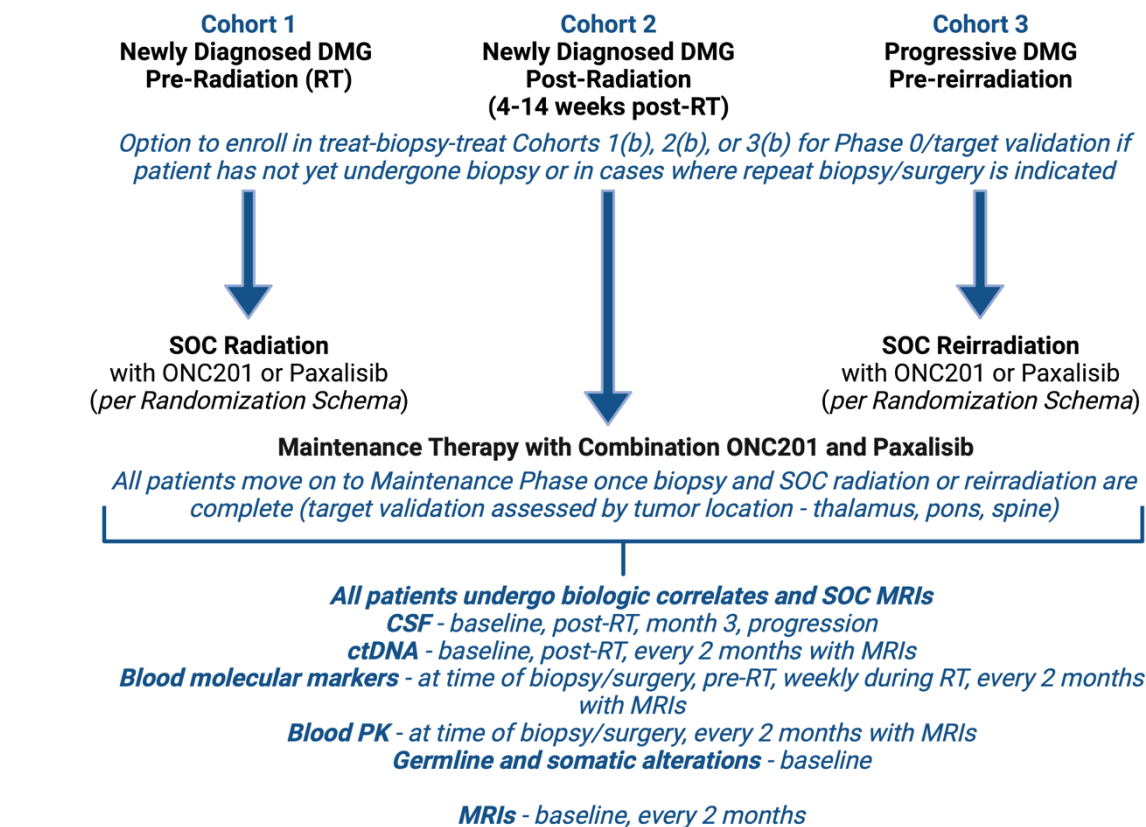
102. Smith EJ, Naik A, Shaffer A, et al. Differentiating radiation necrosis from tumor recurrence: a systematic review and diagnostic meta-analysis comparing imaging modalities. *J Neurooncol.* 2023;162(1):15-23. doi:10.1007/s11060-023-04262-1
103. Verger A, Kas A, Darcourt J, Guedj E. PET Imaging in Neuro-Oncology: An Update and Overview of a Rapidly Growing Area. *Cancers.* 2022;14(5):1103. doi:10.3390/cancers14051103
104. Rosenfeld A, Etzl M, Bandy D, et al. Use of Positron Emission Tomography in the Evaluation of Diffuse Intrinsic Brainstem Gliomas in Children. *J Pediatr Hematol Oncol.* 2011;33(5):369-373. doi:10.1097/MPH.0b013e31820ad915
105. Tinkle CL, Duncan EC, Doubrovin M, et al. Evaluation of ¹¹C-Methionine PET and Anatomic MRI Associations in Diffuse Intrinsic Pontine Glioma. *J Nucl Med.* 2019;60(3):312-319. doi:10.2967/jnumed.118.212514
106. Piccardo A, Tortora D, Mascelli S, et al. Advanced MR imaging and 18F-DOPA PET characteristics of H3K27M-mutant and wild-type pediatric diffuse midline gliomas. *Eur J Nucl Med Mol Imaging.* 2019;46(8):1685-1694. doi:10.1007/s00259-019-04333-4
107. Morana G, Tortora D, Bottoni G, et al. Correlation of multimodal ¹⁸F-DOPA PET and conventional MRI with treatment response and survival in children with diffuse intrinsic pontine gliomas. *Theranostics.* 2020;10(26):11881-11891. doi:10.7150/thno.50598
108. Fiz F, Bini F, Gabriele E, et al. Role of Dynamic Parameters of 18F-DOPA PET/CT in Pediatric Gliomas. *Clin Nucl Med.* 2022;47(6):517-524. doi:10.1097/RLU.0000000000004185
109. Albert NL, Weller M, Suchorska B, et al. Response Assessment in Neuro-Oncology working group and European Association for Neuro-Oncology recommendations for the clinical use of PET imaging in gliomas. *Neuro-Oncol.* 2016;18(9):1199-1208. doi:10.1093/neuonc/nov058
110. Dunkl V, Cleff C, Stoffels G, et al. The Usefulness of Dynamic O-(2-¹⁸F-Fluoroethyl)-l-Tyrosine PET in the Clinical Evaluation of Brain Tumors in Children and Adolescents. *J Nucl Med.* 2015;56(1):88-92. doi:10.2967/jnumed.114.148734
111. Vettermann FJ, Unterrainer M, Ruf V, et al. Dual PET Imaging of an H3K27M-Mutant Glioma With 18F-GE-180 and 18F-FET PET. *Clin Nucl Med.* 2020;45(12):992-993. doi:10.1097/RLU.0000000000003331
112. Marnier L, Lundemann M, Sehested A, et al. Diagnostic accuracy and clinical impact of [18F]FET PET in childhood CNS tumors. *Neuro-Oncol.* 2021;23(12):2107-2116. doi:10.1093/neuonc/noab096

Figure Legend

Figure 1. Schematic of PNOC022. Each cohort (1, 2, 3) is represented at the top of the figure with each phase of treatment within the respective cohort outlined as the figure moves from top to bottom. Description and frequency of collected biologic and imaging correlates on PNOC022 is detailed at the bottom of the figure. CSF, cerebrospinal fluid; ctDNA, circulating tumor DNA; DMG, diffuse midline glioma; MRI, magnetic resonance imaging; PK, pharmacokinetics; RT, radiation therapy. Created with Biorender.com.

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Figure 1



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